

Mesoscopic simulations of lipid membranes

Mingyang Hu, Patrick Diggins, and Markus Deserno

Department of Physics, Carnegie Mellon University, Pittsburgh PA, USA

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Introduction

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Why coarse-graining?

Solvent-free CG Model

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Protein-induced budding

Lipid A-B-mixtures
stretching

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mingyang@cmu.edu

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What's a lipid membrane?

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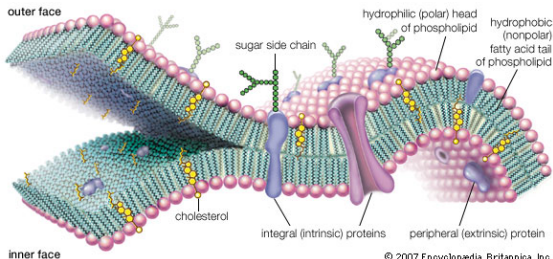
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- ▶ forms continuous barriers around cells, cell nuclei
- ▶ made of two layers of lipid molecules
- ▶ controls the diffusion of molecules in and out of the cell

Building blocks: Lipid molecules

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mingyang@cmu.edu

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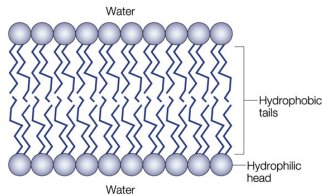
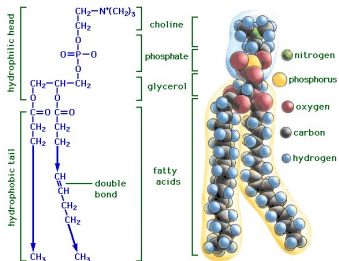
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► amphiphilic molecule

- hydrophilic head (polar: “attracted” by water)
- hydrophobic tails (apolar: “repelled” by water)

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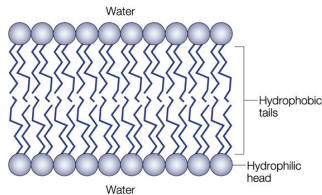
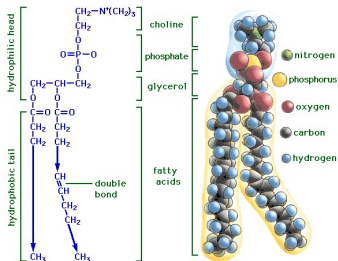
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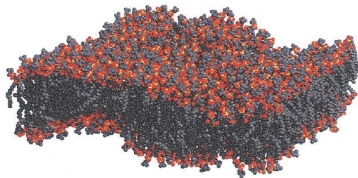


- ▶ amphiphilic molecule
 - ▶ hydrophilic head (polar: “attracted” by water)
 - ▶ hydrophobic tails (apolar: “repelled” by water)
- ▶ membranes form by spontaneous aggregation of lipids
 - ▶ *self-assembly* process

Why coarse-graining?

Lindahl, E. & Edholm, O. *Mesoscopic undulations and thickness fluctuations in lipid bilayers from molecular dynamics simulations*. *Biophys. J.* **79**, 426-433 (2000)

All-atom lipid bilayer
20 nm × 20 nm, 1024 lipids
simulation time: 10 ns



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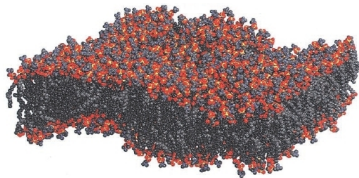
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What if we want a boxlength of $L = 200\text{nm}$?

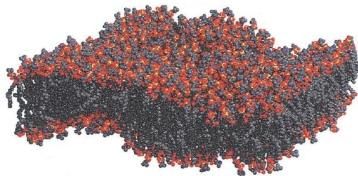
How does computing effort scale with L ?

$$\text{effort} \sim \underbrace{L^2}_{\text{geometry}}$$

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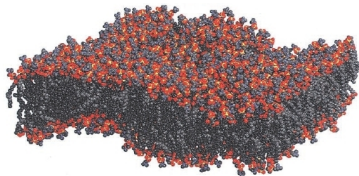
How does computing effort scale with L ?

$$\text{effort} \sim \underbrace{L^2}_{\text{geometry}} \times \underbrace{L^4}_{\text{equilibration}} \sim L^6$$

Efficiency

20 nm → 200 nm

Million times more
computationally expensive!

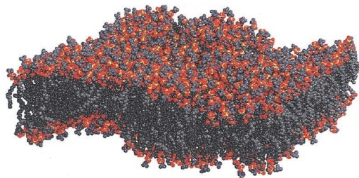


- ▶ amount of material \sim membrane area $A = L^2$.
- ▶ domain decomposition scheme:
 - ▶ increase # CPUs $\sim A$
- ▶ but simulation time $\sim A^3$
 - ▶ uncompensated factor of A^2

Efficiency

20 nm → 200 nm

Million times more
computationally expensive!



- ▶ CG helps to understand the essence of the problems
- ▶ CG reduces the number of DOF
- ▶ CG allows larger time steps
- ▶ CG smoothens the free energy surface
⇒ Better sampling

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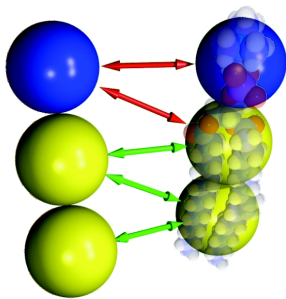
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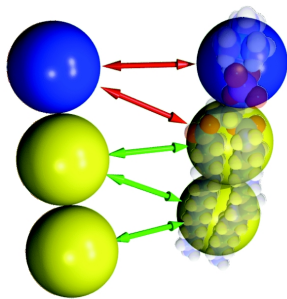
The CG model¹



- Coarse-grain in order to probe the mesoscopic regime of lipid bilayers
- ▶ generic top-down bead-spring
 - ▶ 3 beads: reasonable aspect ratio
 - ▶ only pair forces
 - ▶ solvent free

¹I.R. Cooke, K. Kremer, M. Deserno, Phys. Rev. E **72**, 011506 (2005)
I.R. Cooke and M. Deserno, J. Chem. Phys. **123**, 224710 (2005).

The CG model¹



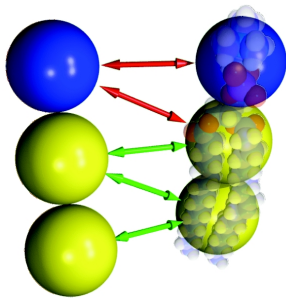
Coarse-grain in order to probe the mesoscopic regime of lipid bilayers

- ▶ generic top-down bead-spring
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- ▶ Note: several other similar models exist.
- ▶ Here I'll talk about the one used in our group
- ▶ It has been implemented in ESPResSo.

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The CG model¹



Coarse-grain in order to probe the mesoscopic regime of lipid bilayers

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Why is “solvent free” good?



membrane



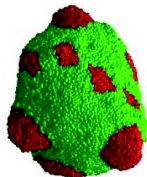
surface



solvent

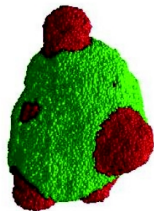


bulk



Example^a

- ▶ 16,000 DPD lipids, 4 beads per lipid.
- ▶ 64,000 particles for lipids.
- ▶ But in total 1,536,000 particles in box!



96% simulation time spent with solvent.

^aM. Laradji & P.B. Sunil Kumar, Phys. Rev. Lett. **93**, 198105 (2004).

Why is “solvent free” good?



membrane



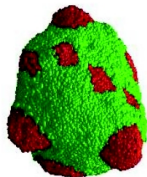
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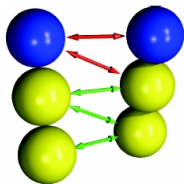


They had a good reason for doing this:
study of the *dynamics* of domain growth,
where hydrodynamics is an important factor.

^aM. Laradji & P.B. Sunil Kumar, Phys. Rev. Lett. **93**, 198105 (2004).

Difficulties

- ▶ Implicit solvent models are incredibly common and useful in polymer physics.
- ▶ Why has it taken so long for them to appear in the field of membrane research?
 - ▶ Polymers don't first have to self assemble!
- ▶ One needs additional cohesion to make the lipids come together.
- ▶ Fluidity has proven to be the major challenge.



Lennard-Jones interactions:

- ▶ weak attraction → gas phase
- ▶ **no fluid phase in between?!**
- ▶ strong attraction → solid (gel) phase

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- ▶ link three beads

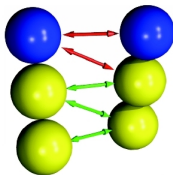
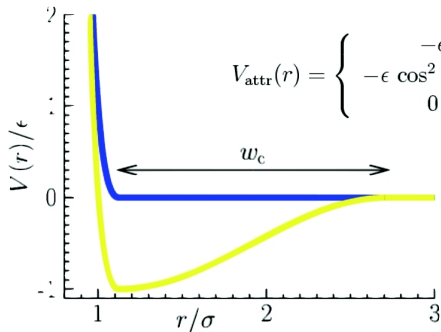
$$V_{\text{bond}}(r) = -\frac{1}{2}k_{\text{bond}}r_{\infty}^2 \ln [1 - (r/r_{\infty}^2)]$$

- ▶ make lipid stiff

$$V_{\text{bend}}(r_{13}) = \frac{1}{2}k_{\text{bend}}(r_{13} - 4\sigma)^2$$

- ▶ nonbonded

$$V_{\text{rep}}(r) = 4\epsilon \left[\left(\frac{r_c}{r}\right)^{12} - \left(\frac{r_c}{r}\right)^6 + \frac{1}{4} \right] \Theta(r_c - r)$$



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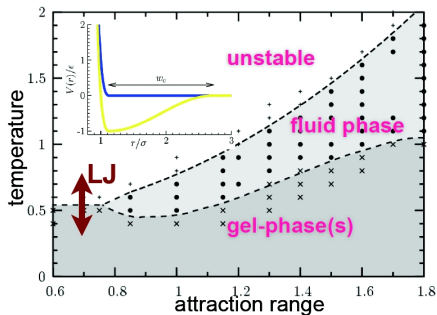
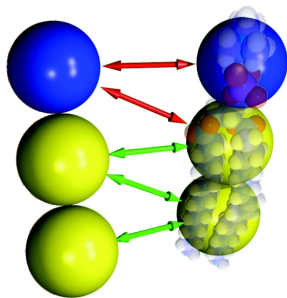
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Overall phase behavior



- ▶ long-range attractions “save” the system some entropy!

Self-assembly

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Mingyang Hu

mingyang@cmu.edu

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`movies/self_assembly_1.mpg`

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- ▶ `mbtools` architecture:
 - ▶ engine (C)
 - ▶ interactions (lj-cos² interaction implemented as a FEATURE)
 - ▶ several analysis routines (e.g. modes, stress tensor)
 - ▶ user-interface (Tcl)
 - ▶ system generation (e.g. initial particle positions)
 - ▶ parameters (configuration files)
 - ▶ register particles in ESPResSo (e.g. topology, interactions)
 - ▶ call to `integrate` command
 - ▶ analysis/output

Program structure located in:

`ESPRESSO/packages/mbtools`

Program structure

- ▶ `mbtools` is a Tcl *package*.
 - ▶ set of Tcl functions
 - ▶ (crude) version control
- ▶ load package to get access to enclosed functions
`package require mbtools [1.0.0]`
- ▶ ESPResSo loads `mbtools` during initialization (file `ESPRESSO/scripts/init.tcl`)
`lappend auto_path "[pwd]/packages/mbtools/"`
- ▶ `auto_path` contains the list of (sub)directories checked by the package loader
- ▶ use *namespaces* to avoid defining two functions with the same name

Program structure

```
# Espresso/packages/mbtools/mbtools.tcl
#

package require ::mbtools::utils
package require ::mbtools::system_generation
package require ::mbtools::analysis

package provide mbtools 1.0.0

namespace eval mbtools {

}

# Espresso/packages/mbtools/utils/setup.tcl
#

proc ::mbtools::utils::readcheckpoint { dir }
```

Config file example (incomplete...)

```
# .../mbtools/examples/simplebilayer.tcl
#

# define geometry
set geometry { geometry "flat -fixz" }
# time step
set main_time_step 0.01
# analysis
lappend analysis_flags pressure
```

From command line:

```
Espresso scripts/main.tcl simplebilayer.tcl
```

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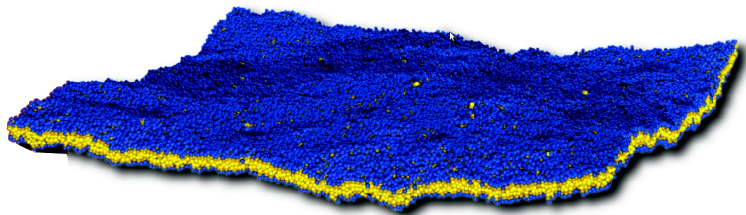
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Membrane elasticity

- ▶ Model membrane as a 2D elastic sheet (continuum theory)

$$E = \int dA \left\{ \frac{1}{2} \kappa K^2 + \sigma \right\} \simeq \frac{1}{2} \int dx dy \left\{ \kappa (\nabla^2 h)^2 + \sigma (\nabla h)^2 \right\}$$

- ▶ κ : bending modulus
- ▶ K : total curvature
- ▶ σ : surface tension
- ▶ $h(x, y)$: height function (Monge gauge)



Bending modulus

- ▶ Model membrane as a 2D elastic sheet (continuum theory)

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- ▶ κ : bending modulus
 - ▶ K : total curvature
 - ▶ σ : surface tension
 - ▶ $h(x, y)$: height function (Monge gauge)
- ▶ Fourier expansion and equipartition theorem

$$\langle |h_{\mathbf{q}}|^2 \rangle = \frac{k_B T}{L^2 [\kappa q^4 + \underbrace{\sigma q^2}_{\text{set to 0}}]} = \frac{k_B T}{L^2 \kappa} q^{-4}$$

- ▶ determine bending modulus κ

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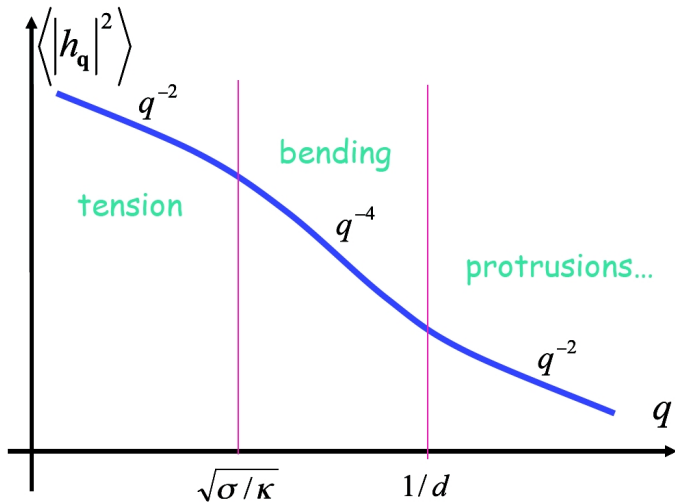
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Fluctuation spectrum from continuum theory

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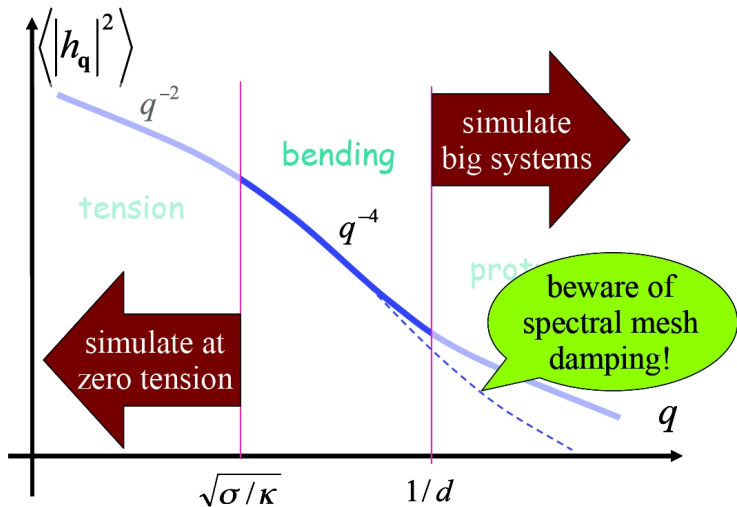
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However...

- ▶ Equilibration time of Fourier modes scales like q^{-4}
- ▶ Large bending modulus κ from small perturbation ($k_B T$)
→ small signal!

$$h(x) = h_q e^{iqx} \quad \rightarrow \quad K = -h''(x) = h_q q^2 e^{iqx}$$

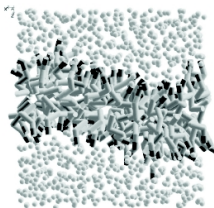
$$\langle K^2 \rangle = \langle |h''(x)|^2 \rangle = q^4 \langle |h_q|^2 \rangle = \frac{k_B T}{L^2 \kappa}$$

$$\bar{R} = \frac{1}{\langle K^2 \rangle^{1/2}} = \sqrt{\frac{\kappa}{k_B T}} L \simeq 3 \dots 5L$$

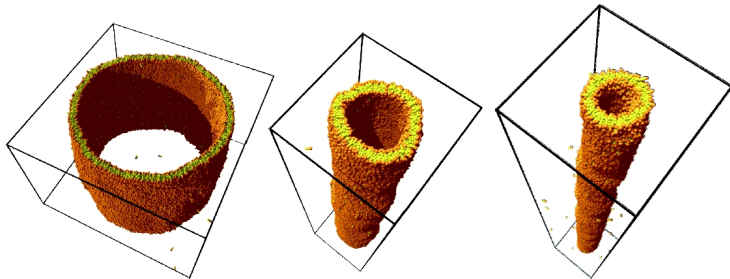
- ▶ Result relevant for strong bending?

κ from actively bent membranes

- ▶ first implementation:
W.K. den Otter and W.J. Briels, J. Chem. Phys. **118**, 4712 (2003)
- ▶ Enforce large undulation mode, measure constraining force.



Simpler way: stretch a membrane tether!²



²V. A. Harmandaris and M. Deserno, J. Chem. Phys. **125**, 204905 (2006)

κ from actively bent membranes³

- ▶ Area:

$$A = 2\pi RL$$

- ▶ Energy:

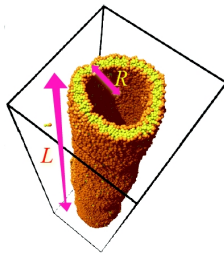
$$E = \frac{\kappa}{2} \times \frac{1}{R^2} \times A$$

- ▶ Force:

$$F = \left(\frac{\partial E}{\partial L} \right)_A = \dots = \frac{2\pi\kappa}{R}$$

- ▶ Bending modulus:

$$\kappa = \frac{FR}{2\pi} \simeq \frac{\bar{F}\bar{R}}{2\pi}$$



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κ from actively bent membranes⁴

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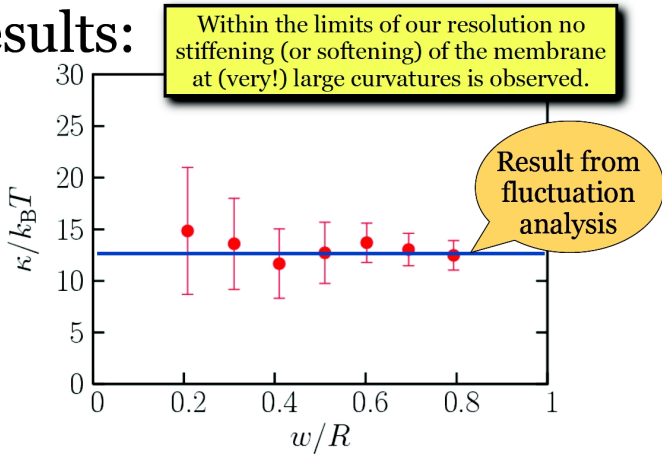
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Results:



⁴V. A. Harmandaris and M. Deserno, J. Chem. Phys. **125**, 204905 (2006)

Simpler way of extracting the line tension

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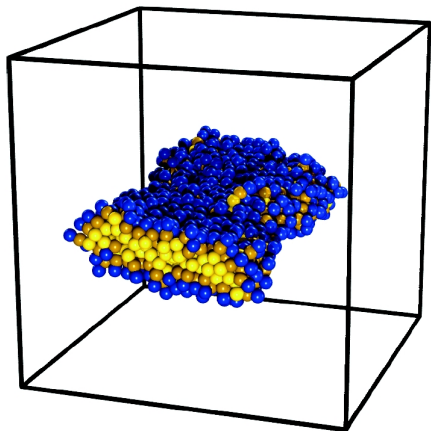
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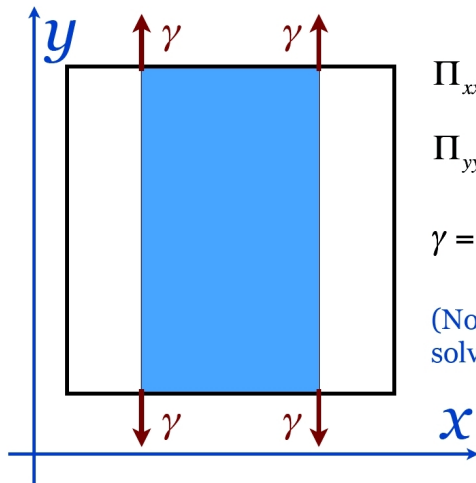
Protein-induced budding

Lipid A-B-mixtures
stretching



- ▶ simulate a periodically half-connected bilayer in a box
- ▶ stress tensor will be imbalanced precisely by twice the line tension!

Simpler way of extracting the line tension



$$\Pi_{xx} = P$$

$$\Pi_{yy} = P - 2\gamma/L_x L_z$$

$$\gamma = \frac{1}{2} L_x L_z (\Pi_{xx} - \Pi_{yy})$$

(Notice that $P = 0$ in the solvent free case!)

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Mingyang Hu

mingyang@cmu.edu

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- ▶ competition between bending rigidity and line tension ⁵
- ▶ sonicate vesicle solution: rip vesicles into bits and pieces!
- ▶ these (flat) pieces will merge and grow bigger
- ▶ at what point will they again close up and form vesicles?

⁵“The size of bilayer vesicles generated by sonication”, W. Helfrich, Physics Letters A, Vol 50, Issue 2, p. 115-116

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Mingyang Hu

mingyang@cmu.edu

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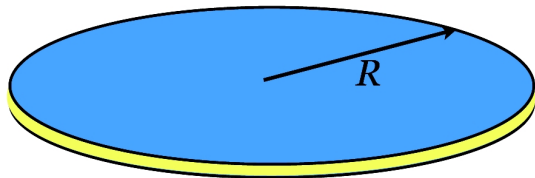
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`movies/c16sa.mpg`

Vesicles



$$E_{\text{pancake}} = 2\pi R \gamma$$

$$E_{\text{vesicle}} = 4\pi(2\kappa + \bar{\kappa})$$

Energies are equal, if

$$R_{\text{pancake}} = \frac{2(2\kappa + \bar{\kappa})}{\gamma}$$

(real stability analysis: 2 \rightarrow 4)

$$\begin{aligned} E_{\text{vesicle}} &= 4\pi R^2 \cdot \left[\frac{1}{2}\kappa \left(\frac{1}{R} + \frac{1}{R} \right)^2 + \bar{\kappa} \frac{1}{R} \cdot \frac{1}{R} \right] \\ &= 4\pi(2\kappa + \bar{\kappa}) \end{aligned}$$

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What values do we expect?

- ▶ $\kappa = 20 k_B T \simeq 80 \text{ pN} \cdot \text{nm}$
- ▶ $\bar{\kappa} \simeq -\kappa$ (very little is known about $\bar{\kappa}$, come back tomorrow!)
- ▶ $\gamma \simeq 10 \text{ pN}$

$$\text{▶ } R_{\text{pancake}} = \frac{4(2\kappa + \bar{\kappa})}{\gamma} \simeq \frac{4\kappa}{\gamma} \simeq \frac{320 \text{ pNnm}}{10 \text{ pN}} = 32 \text{ nm}$$

- ▶ This is then also the diameter of vesicles we expect to find!

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Mingyang Hu

mingyang@cmu.edu

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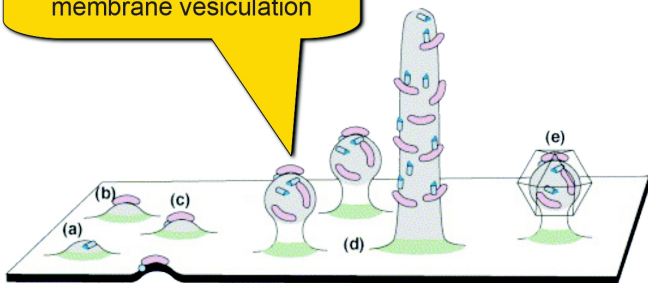
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Membrane-curving proteins
can attract and drive
membrane vesiculation

B. Antony, *Curr. Opin.
Cell Biol.* **18**, 386 (2006)



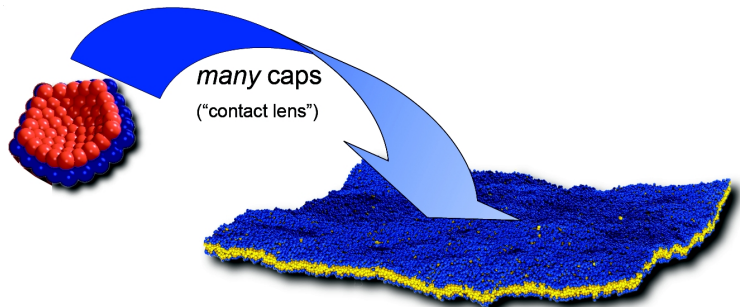
Intuitive, but no physical justification!

Protein-induced budding⁶

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Mingyang Hu

mingyang@cmu.edu



- ▶ 36 curved caps, $\sim 50,000$ lipids
- ▶ 160nm side-length, total time ~ 1 ms
- ▶ no lateral tension
- ▶ no explicit interaction between caps

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⁶B.J. Reynwar et al., Nature **447**, 461 (2007)

Protein-induced budding⁷

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Mingyang Hu

mingyang@cmu.edu

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movies/caps.avi

⁷B.J. Reynwar et al., Nature **447**, 461 (2007)

Protein-induced budding⁸

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Mingyang Hu

mingyang@cmu.edu

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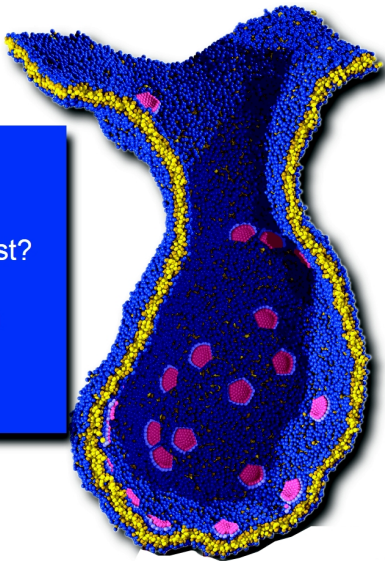
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Some observations:

- Caps **attract** collectively
- Attractive pair-forces exist?
- No crystalline structure
- Cooperative vesiculation
- No “scaffolding”
- 50-100nm length scales
- several milliseconds



⁸B.J. Reynwar et al., Nature **447**, 461 (2007)

Protein-induced budding⁹

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Mingyang Hu

mingyang@cmu.edu

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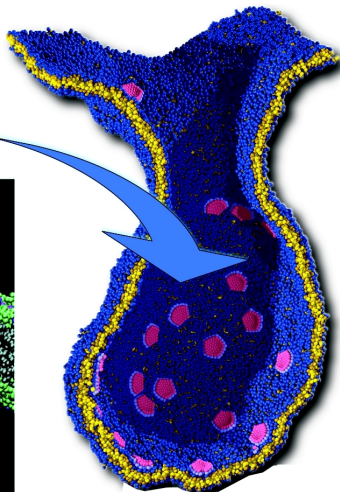
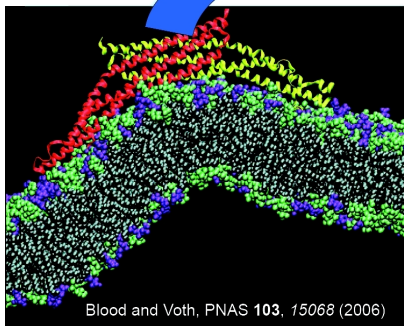
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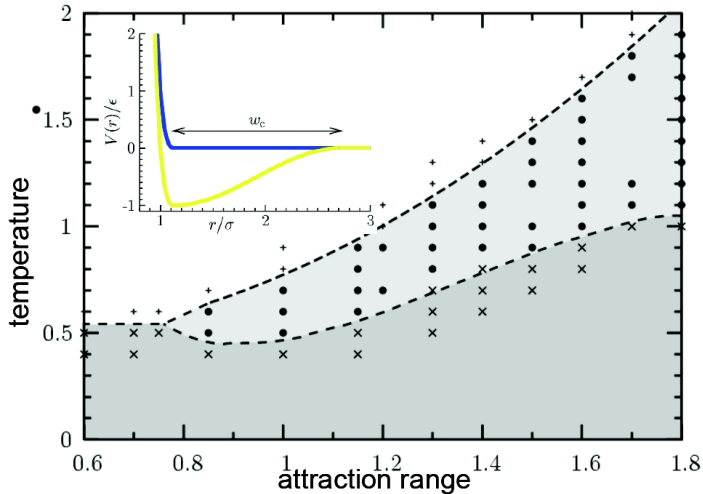
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⁹B.J. Reynwar et al., Nature **447**, 461 (2007)

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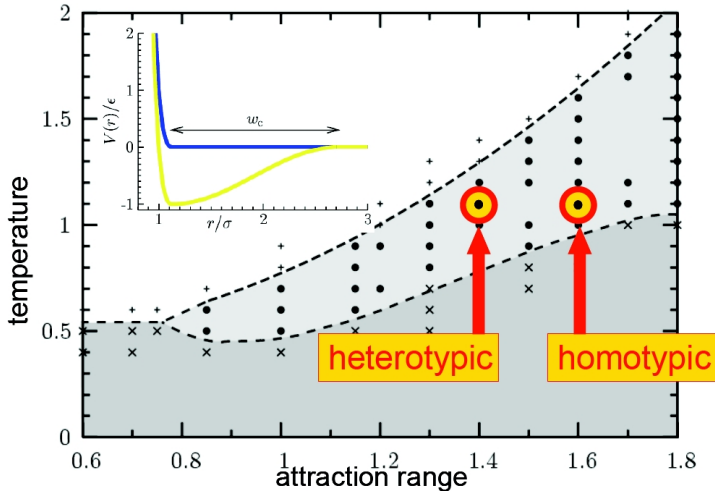
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Lipid A-B-mixtures



$$w_{AB} < w_{AA} = w_{BB}$$

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Lipid A-B-mixtures¹⁰

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Mingyang Hu

mingyang@cmu.edu

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movies/budding.mpg

¹⁰B.J. Reynwar & M. Deserno, Biointerphases **3**, FA118 (2009)

Lipid A-B-mixtures¹¹

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Mingyang Hu

mingyang@cmu.edu

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movies/spinodal.avi

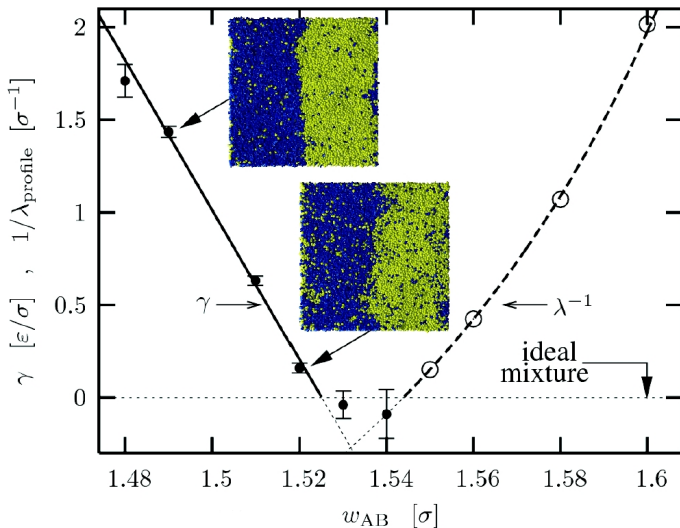
¹¹from Sarah Veatch

Lipid A-B-mixtures

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Mingyang Hu

mingyang@cmu.edu



$$w_{AB} < w_{AA} = w_{BB}$$

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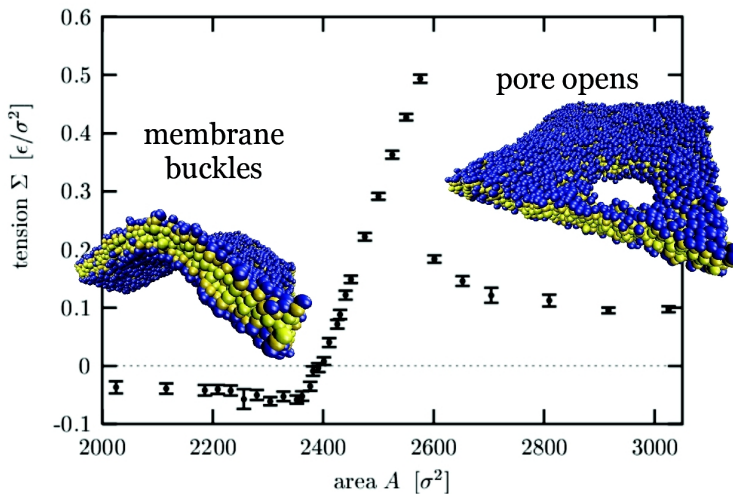
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stretching modulus¹²

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Mingyang Hu

mingyang@cmu.edu



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stretching modulus

simple theory:

farago, jcp, 2003; tolpekina/den otter/briels, jcp 2004; cooke/deserno, jcp 2005

membrane stretching plus line energy

$$e = \frac{1}{2}m \frac{(a - a_s - \pi r^2)^2}{a_s} + 2\pi\gamma r$$

rescaling of energy

$$\lambda^3 = \frac{\gamma a_s}{\pi m}, \quad \tilde{r} = \frac{r}{\lambda}, \quad b = \frac{a - a_s}{\pi \lambda^2}$$

equilibrium condition for pore radius

$$\tilde{r}^3 - b\tilde{r} + 1 = 0$$

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